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# Physiological neural representation for personalised tracer kinetic parameter estimation from dynamic PET

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Abstract. Dynamic positron emission tomography (PET) with tracer [18F]FDG enables non-invasive quantification of glucose metabolism by means of kinetic analysis, often modelled by the two-tissue compartment model (TCKM). However, voxel-wise kinetic parameter estimation using conventional methods is computationally intensive and limited by spatial resolution. Deep neural networks (DNNs) offer an alternative but require large training datasets and significant computational resources. To address these limitations, we propose a physiological neural representation based on implicit neural representations (INRs) for personalized kinetic parameter estimation. INRs, which learn continuous functions, allow for efficient, high-resolution parametric imaging with reduced data requirements. Our method also integrates anatomical priors from a 3D CT foundation model to enhance robustness and precision in kinetic modelling. We evaluate our approach on an [18F]FDG dynamic PET/CT dataset and compare it to state-of-the-art DNNs. Results demonstrate superior spatial resolution, lower mean-squared error, and improved anatomical consistency, particularly in tumour and highly vascularized regions. Our findings highlight the potential of INRs for personalized, data-efficient tracer kinetic modelling, enabling applications in tumour characterization, segmentation, and prognostic assessment. The code is available at: https://github.com/tkartikay/PhysNRPET

**Keywords:** Implicit Neural Representations  $\cdot$  Tracer Kinetic Modelling  $\cdot$  Dynamic PET.

## 1 Introduction

Dynamic positron emission tomography (PET) with [ $^{18}$ F]FDG has allowed for non-invasive assessment of glucose metabolism in tissues through kinetic analysis, in particular with the irreversible two-compartment-kinetic model (TCKM). The kinetic parameters thus obtained,  $K_1$ ,  $k_2$ , and  $k_3$  represent the rates of

transport of the tracer between blood and cells (influx and efflux) and the rate of phosphorylation of glucose, respectively.  $V_B$  describes the blood volume fraction, to correct for the partial volume effects arising from the limitations of the scanner spatial resolutions [6]. Since glucose metabolism differs for metastatic lesions from non-metastatic tissues with increased uptake, kinetic parameters have been advantageous in successful diagnosis, complementing Standard Uptake Values (SUV) values from static PET images [22]. Phosphorylation rate  $(k_3)$  is significant metric for tumour differentiation in colorectal cancers [19] and breast cancers [8], and the perfusion parameter  $(K_1)$  can be a marker of angiogenesis in breast cancers [3]. Kinetic parameters, carrying more information than SUV, can play a vital role as additional biomarkers for tumour heterogeneity analysis, precise segmentations, and prognostic predictions.

Conventional methods based on non-linear curve fitting (CF) used to perform kinetic analysis on volumes of interest lack the spatial neighbourhood understanding and are slow when applied to derive voxel-based parametric images. These challenges can be overcome through the use of deep neural networks (DNNs) to derive the parametric images [4,7,9]. Self-supervised DNNs have recently been proposed to generate the  $K_1, k_2, k_3, V_B$  parametric images which satisfy the TCKM [5]. These methods, in turn, require large amount of training data to achieve accurate results, and while they are quick to predict the outputs once trained, the training of these networks itself is a slow and memory-intensive process, balanced by reducing the resolution of the training dataset, and slicing it into 2D+t data instead of processing the entire 3D+t data at once.

Implicit neural representations (INRs) are neural networks that learn continuous functions, which can parametrise signals, such as mapping continuously the image intensities from the input spatial coordinates. With steady adoptions in the medical imaging field due to their memory efficiency, improved reconstructions, interpolation capabilities, and personalisation [10], INRs could overcome the data limitation issues faced by DNNs and speed issues of CF in generation of parametric images in dynamic PET modelling. Since INRs can be optimised on single patient acquisitions, they can be adapted to individual patient anatomy and physiology [17], leading to personalised neural networks.

Encoding the dynamic physiological processes in the INR also allows their integration with downstream models that are trained for classification of malignancies, or early detection of metastases, etc., as well as with foundation models that extract features from the associated anatomical data such as CT scans, enabling feature-rich end-to-end pipelines for aiding the clinical routines.

Thus, guided by their promise, we propose the use of INRs for modelling dynamic PET to generate kinetic parameters (Fig. 1). Our contribution is:

- the introduction of a physiological neural representation for tracer kinetic modelling for a [<sup>18</sup>F]FDG dynamic PET/CT dataset,
- its extension by integrating anatomical information and features extracted from corresponding CT scan using a 3D CT foundation model, and
- a comparative analysis against state-of-the-art DNNs and CF in terms of goodness of fit, memory and speed.

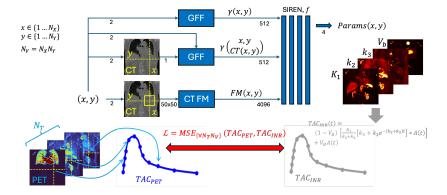


Fig. 1. Schematic overview of the proposed pipeline (illustrated in 2D for clarity; works in 3D spatial coordinates). Spatial positions are encoded using either positional coordinates alone, with local CT Hounsfield units, or with neighbourhood features from a CT foundation model. These features are mapped via a SIREN-based INR to estimate voxelwise kinetic parameters of the irreversible two-tissue compartment model. Predicted parameters generate modelled time-activity curves (TAC), which are optimised by minimising the MSE with measured PET curves.

## 1.1 Related Works

Voxel-wise estimation of kinetic parameters have been explored by solving the non-linear optimisation problem of fitting the model-predicted curves onto the measured data [1,21]. The data in these studies was limited to smallest frames of 10s resolution. With the advancements in long axial field of view scanners, and the availability of improved temporal resolution, the kinetic parameters have also been estimated for increased resolution of 2s frames [14]. Sari et al. provide reference values for kinetic parameters for volumes of interest in oncological patients, and have also shown the voxel-wise generation of TCKM. Building upon this work, de Benetti el al. proposed DNNs for the generation of parametric images from the dynamic PET data, relying on self-supervised training of the DNN through the integration of the kinetic model into the loss function [5]. To parametrise tracer kinetics through encoding the PET reconstruction and one compartment model in neural networks, a hybrid physics- and data-driven approaches is proposed to incorporate the mathematical expressions defining the kinetics [23]. Shao et al. propose employing DNNs to discretise the data, and solving the ODEs that define the kinetic model, where the tracer concentration in each compartment is represented with separate fully connected DNNs, and the kinetic parameters are optimised while minimising the ODE loss [15].

Since INRs are based on the assumption of continuity as an *a priori* [10], these representations can be made robust to noise and further constrained anatomically through image-derived priors, such as extracted from large-scale "foundation" models. Large-scale models trained on high-resolution oncological data can provide knowledge of vascular architecture, tissue heterogeneity, and image-

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based biomarkers as embeddings or input features to the task-specific models. Neural networks that encode the tracer kinetics could similarly benefit from the anatomical understanding of macroscopic blood supply routes, sharp gradients marking organ bounds, regions of tissue similarities and anomalies, etc. A foundation model trained on high-resolution 3D CT oncological datasets with understanding of underlying biology, and shown to improve performance of downstream tasks, particularly with limited dataset, and resilient to input variation has been presented by Pai et al [11].

# 2 Materials and Methods

#### 2.1 Dataset

The dataset consists of 24 patients with various tumour types undergoing dynamic PET imaging (average injected activity:  $235 \pm 51$  MBq of [ $^{18}$ F]FDG), immediately followed by a CT scan using a Siemens Biograph Vision Quadra long-field-of-view PET/CT device. The PET acquisition lasted 65 minutes for all patients and consisted of 62 frames with varying durations: 2 seconds for the initial frames, 30 seconds after the first 2 minutes, and up to 5 minutes per frame toward the end of the acquisition. PET images were reconstructed with a voxel size of  $1.65 \times 1.65 \times 1.65$  mm³ and subsequently filtered using a 2 mm full-width at half maximum (FWHM) Gaussian filter to generate **high-resolution images (HiRes)**. To match the resolution used by De Benetti et al., we down-sampled the PET images to  $2.5 \times 2.5 \times 2.5$  mm³, producing **low-resolution images (LoRes)**. The CT scans were acquired in low-dose mode (voltage: 120 kV, tube current: 25 mA) and reconstructed with a voxel size matching that of the high-resolution PET images.

### 2.2 Preprocessing and Architecture

Dynamic PET data were first normalised to the range [0,1] by dividing by the maximum of the image-derived input function, which was on the order of  $\sim 200~\mathrm{kBq/ml}$ . The corresponding CT data were similarly mapped from [-1024, 2048] to the interval [0,1]. Registration of PET and CT was assumed valid and no motion correction between frames was applied.

Using the normalised CT volume, the CT foundation model [11] extracted 4096 features for each voxel, by taking as input  $50 \times 50 \times 50$  neighbourhood centred on that voxel. These features served as an optional input to the network, depending on the variant of the INR under consideration.

Spatial coordinates  $x \in \mathbb{R}^3$  were first normalised to the interval [-1,1]. Each normalised coordinate was encoded using Gaussian Fourier Features (GFF) [20] [12] drawn from a zero-mean Gaussian distribution, with a standard deviation chosen to control frequency bandwidth. A total of 256 frequencies were sampled, and a multiplier of 10 was applied to spread the frequency spectrum. Specifically, for each coordinate x, the encoding was formed by computing:

 $\gamma(\boldsymbol{x}) = [\sin(2\pi \boldsymbol{B}\boldsymbol{x}),\cos(2\pi \boldsymbol{B}\boldsymbol{x})]^T$  where  $\boldsymbol{B}$  is a randomly generated matrix of size  $256 \times 3$ . The choice of 256 frequencies and a multiplier of 10 followed typical recommendations in the literature and was not varied [20]. In all cases, the spatial domain (2D or 3D) was sampled, and the resulting coordinates were normalised to the interval [-1,1] prior to being processed with GFF. Where relevant (CTHU variant, see below), the voxel-wise H.U. value, also normalised to [0,1], was appended to the normalised coordinates to form an extended input vector. This augmented input was then encoded with GFF. However, in the variant utilising the foundation model features (CTFM variant, see below), the 4096-dimensional feature vector was incorporated directly, without GFF encoding, while GFF was still applied to the spatial coordinates.

The encoded features were passed to a SIREN [18] network with three hidden layers. Each hidden layer comprised 512 units and used sine activations, following the standard weight-initialisation scheme described in the SIREN work to preserve gradient stability. The final layer was a linear layer producing four outputs, the predicted kinetic parameters:  $K_1$ ,  $k_2$ ,  $k_3$ , and  $V_b$ . These outputs correspond to the parameters in the TCKM governed by a system of ordinary differential equations (ODEs). Maintaining differentiability from inputs to outputs facilitated direct gradient-based training in conjunction with the ODE formulation.

Parameter estimation was performed by minimising the mean-squared error (MSE) between the Time Activity Curve (TAC) generated from the predicted parameters and real TAC values measured at the input coordinate in the dynamic PET. Training used the Adam optimiser with a learning rate of  $1e^{-5}$ . The network was trained for 100 epochs and all hyperparameters, including the number of frequencies, the sine activation design, and the network depth, were adapted from SIREN and GFF examples without additional ablation experiments[2,13,16].

Using the estimated kinetic parameters from the INR for each voxel coordinate, the predicted TAC was generated by solving the TCKM ODEs (Eq. 2.2).

$$TAC_{INR}(t) = (1 - V_b) \left[ \frac{K_1}{k_2 + k_3} \left[ k_3 + k_2 e^{-(k_2 + k_3)t} \right] * A(t) \right] + V_b A(t)$$
 (1)

To train the INR, the predicted TAC was then compared using MSE against the observed TAC at the discrete time points where measurements were available. The mean and standard deviation of these reconstruction errors were computed across the full 3D spatial domain with time (3D+t), selected 2D slices with time (2D+t), and at each individual voxel with time (voxel+t). The blood-to-tissue transport constant  $K_i$  was calculated using  $K_i = K_1k_3/(k_2 + k_3)$ .

## 3 Results

We evaluated the proposed architecture in several variants: HiRes and LoRes, 2D (single slice) and 3D (a stack of 10 adjacent slices) - resulting in the variants

inr-HiRes-2D, inr-LoRes-2D, inr-HiRes-3D and inr-LoRes-3D. Due to incomplete data of one patient, we compared inr-HiRes-2D to conventional CF in 23 patients (Tab. 1). In those 23 patients, we manually segmented liver, spleen, both lungs, and both kidneys for analysis of kinetic parameters (Fig. 2). Additionally, we tested configurations incorporating the Hounsfield unit (HU) values from the CT scan (only for HiRes and in 2D - variant CTHU-2D) and those using features extracted with the foundation model (variant CTFM-2D), comparing them to the neural network proposed by De Benetti et al. in both 2D (single-slice) and 3D (stack of 10 adjacent slices) configurations (Tab. 2) [5].

As comparison metrics we used MSE with its standard deviation (SD), and training time, inference time, and memory utilization. All experiments were conducted on a MacBook with an M3 Pro (12-core CPU, Metal 3, 36GB RAM).

**Table 1.** Quantitative evaluation of INR against CF: differences are statistically significant  $(p \ll 0.05)$ 

Method	Mean MSE ( $\pm$ SD)	SD MSE	Runtime (± SD)
INR	$5.48 ext{E-}03 \pm 2.43 ext{E-}03$	1.13E-02	$1336.13\pm38.54\;\mathrm{s}$
CF	$1.16\text{E}$ -02 $\pm$ 8.23E-03	2.27E-02	$40933.44 \pm 1651.57 \text{ s}$

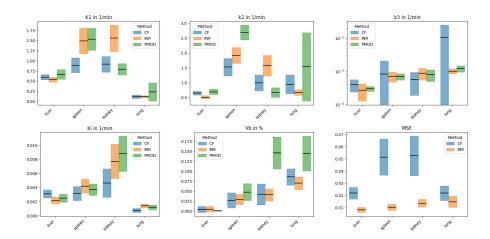
**Table 2.** Quantitative evaluation of INR variants and comparison with the spatio-temporal neural network of De Benetti et al. (FDB112-2D and FDB112-3D).

Model	Voxel	$MSE(\pm SD)$	Training	Inference	Memory
	Resolution		Time	Time	Utilisation
FDB112-3D	2.5 mm	$0.066 \pm 0.255$	-	62 s	-
INR-LoRes-3D	2.5  mm	$0.009 \pm 0.093$	24.3 min	66 s	6.7 GB
FDB112-2D	2.5 mm	$0.073 \pm 0.268$	-	$0.15 \; { m s}$	-
INR-LoRes-2D	2.5  mm	$0.009 \pm 0.093$	24.1 min	6 s	4.3 GB
INR-2D-HiRes	1.65 mm	$0.009 \pm 0.094$	25.2 min	8 s	11.9 GB
CTHU-INR-2D-HiRes	1.65  mm	$0.009 \pm 0.094$	25.4 min	8 s	$12.4~\mathrm{GB}$
CTFM-INR-2D-HiRes	1.65  mm	$0.009 \pm 0.094$	$25.7 \min$	9 s	13 GB

To qualitatively visualize the impact of the different INR variants, we visualize the mean MSE and its SD (Fig. 3) for an exemplary patient. This gives an idea of which organs the measured and the calculated TACs depart in average (mean MSE), and its spread (SD) is. Additionally, Fig. 4 shows for a particular segment across the tumour when in time and space the MSE is highest.

# 4 Discussion

This study applies implicit neural representations (INRs) to tracer kinetic modelling in dynamic [<sup>18</sup>F]FDG PET. Unlike non-linear CF or large-scale DNNs,



**Fig. 2.** Box plots (average  $\pm$  SD for calculated parameters and MSE for four organs across 23 patients using INR and CF. Ranges obtained with PMOD as reported by Sari et al. [14] are plotted alongside for reference.

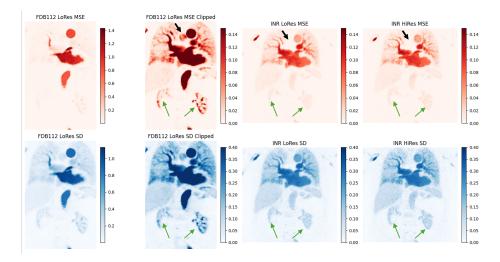
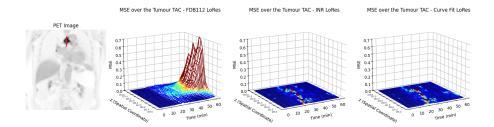


Fig. 3. MSE (top row, red) and SD (lower row, blue) for a coronal slice of an exemplary patient. MMSE is higher on average over the complete slice (see FDB112 LoRes MSE Clipped vs. INR LoRes MSE), particularly in the tumour (black arrow) and kidneys (green arrow), ignoring the blood pool (visible larger vessels). SD images show less variation on the MSE over time for our model. LoRes and HiRes variants of our model (right two columns) show good agreement with more details in the HiRes image.



**Fig. 4.** MSE along a segment crossing a mediastinal lymph node metastasis (see PET slice, left). Plots show MSE as a function of space (left to bottom axis) and time (bottom to right axis). The left plot displays the 2D baseline; the centre plot shows the inr-LoRes-2D variant; the right plot shows the curve fit results. The baseline exhibits higher error in the lesion centre at late time points, whereas the INR yields lower MSE similar to CF, with errors instead aligning with the IDIF peak (not shown).

INRs approximate the continuous spatio-temporal function of patient-specific PET signals. Such a representation requires fewer training examples than conventional DNNs, making it useful when data availability is limited. When compared to CF, calculating dynamic parameters with INRs takes significantly shorter time and produces smoother images as spatial information is integrated regularizing the noise at a single voxel level. In addition, modelling kinetics through an INR allows more flexibility in handling anatomical and physiological variability across patients.

We integrated anatomical information from CT by incorporating HU values and, in a separate variant, 3D CT foundation model features. Both approaches provided comparable MSE but showed slightly faster convergence (not shown), suggesting that anatomical descriptors can guide the estimation of kinetic parameters. The proposed INRs consistently achieved lower MSE than a baseline DNN and CF, likely due to their ability to capture spatio-temporal dynamics via a continuous, data-efficient mapping.

Performance across INR variants was similar in accuracy, though convergence speed and memory demands differed. Foundation-model-based INRs converged faster but required more memory. Nonetheless, LoRes variants of our INR reduced the resource footprint, enabling training on standard hardware setups.

Comparisons with a baseline DNN and CF to a lesser extent proved that INRs show lower errors, especially in tumour and kidney regions, which often exhibit rapid or heterogeneous tracer kinetics. This highlights the robust parameter estimation of INRs when organ- or lesion-specific kinetic variability must be captured.

Accurate kinetic parameters  $(K_i, K_1, k_2, k_3, V_b)$  can reveal metabolic properties that aid in tumour assessment and therapy evaluation. The continuous form of INRs and their integration with anatomical descriptors can potentially reduce segmentation errors and support downstream tasks such as lesion clas-

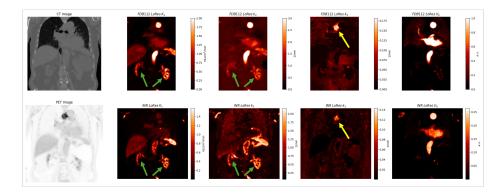


Fig. 5. Coronal slices of parametric images from the 2D baseline (upper row, second to last columns) and the inr-LoRes-2D variant. INR-derived maps exhibit greater detail and sharper edges. Boundaries of the kidneys (green arrow, prominent in  $K_1$  and  $k_2$ ) and tumour (yellow arrow, seen in  $k_3$ ) are more clearly defined, highlighting tissue heterogeneity. The higher contrast in the tumour indicates greater specificity.

sification. Further research could expand the scope by incorporating additional imaging modalities, testing on larger datasets, or refining network architectures and Fourier feature encodings. This future work may improve accuracy, promote generalisability, and further lower the hardware demands for advanced tracer kinetic modelling.

**Disclosure of Interests.** The authors have no competing interests to declare that are relevant to the content of this article.

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